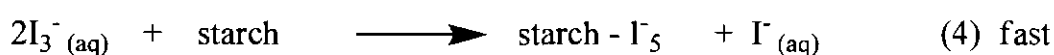
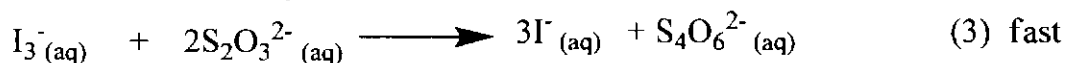
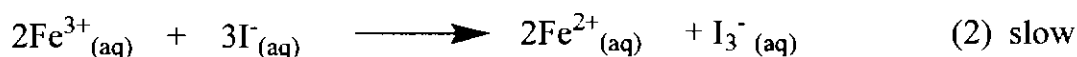


PRACTICAL EXAMINATION

Practical Problem 1 14 % of the total	Code:		Question	1	2	3	4	5	6	Total
	Examiner		Mark	2	4	50	2	2	10	70
	Grade									

Practical Problem 1. The oxidation of iodide by iron(III) ions – a kinetic study based on the thiosulfate clock reaction

Clock reactions are commonly used as demonstrations by chemical educators owing to their visual appeal. Oxidation of iodide by iron(III) ions in a weakly acidic medium is a reaction that can be transformed into a clock reaction. In the presence of thiosulfate and starch, chemical changes in this clock reaction can be presented by the following equations:



Reaction (1) is a fast reversible equilibrium which occurs in the reaction mixture giving a reservoir of iron(III) and thiosulfate ions. After being produced in reaction (2), iodine in the form of triiodide ion (I_3^-), is immediately consumed by thiosulfate in reaction (3). Therefore, no iodine accumulates in the presence of thiosulfate. When thiosulfate is totally depleted, the triiodide ion accumulates and it may be detected by use of starch indicator according to reaction (4).

The kinetics of reaction (2) is easily investigated using the initial rates method. One has to measure the time elapsed between mixing the two solutions and the sudden color change.

For the oxidation of iodide by iron(III) ions (reaction 2), the reaction rate can be defined as:

$$v = -\frac{d[\text{Fe}^{3+}]}{dt} \quad (5)$$



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The initial reaction rate can then be approximated by:

$$v_0 \approx -\frac{\Delta[\text{Fe}^{3+}]}{\Delta t} \quad (6)$$

with $\Delta[\text{Fe}^{3+}]$ being the change in the concentration of iron(III) ions in the initial period of the reaction. If Δt is the time measured, then $\Delta[\text{Fe}^{3+}]$ is the change in iron(III) ion concentration from the moment of mixing to the moment of complete thiosulfate consumption (assume that the reaction rate does not depend on thiosulfate concentration). Therefore, from the reactions' stoichiometry it follows:

$$-\Delta[\text{Fe}^{3+}] = [\text{S}_2\text{O}_3^{2-}]_0 \quad (7)$$

and consequently:

$$v_0 \approx \frac{[\text{S}_2\text{O}_3^{2-}]_0}{\Delta t} \quad (8)$$

The initial thiosulfate concentration is constant and significantly lower than that of iron(III) and iodide ions. The above expression enables us to determine the initial reaction rate by measuring the time required for the sudden color change to take place, Δt .

The rate of reaction is first order with respect to $[\text{Fe}^{3+}]$, and you will determine the order with respect to $[\text{I}^-]$. This means the initial reaction rate of reaction can be expressed as:

$$v_0 = k[\text{Fe}^{3+}]_0 [\text{I}^-]_0^y \quad (9)$$

where k is the rate constant and y is the order with respect to $[\text{I}^-]$.

We assume that the reaction rate does not depend on the thiosulfate concentration, and that the reaction between Fe^{3+} and $\text{S}_2\text{O}_3^{2-}$ is negligible. You have to observe carefully the color changes during the clock reaction and to determine the reaction order with respect to $[\text{I}^-]$, and the rate constant of clock reaction.

Experimental Set-up

Instructions for using the digital timer (stopwatch)

1. Press the [MODE] button until the 00:00:00 icon is displayed.
2. To begin timing, press the [START/STOP] button.
3. To stop timing, press the [START/STOP] button again.
4. To clear the display, press the [SPLIT/RESET] button.

PRECAUTIONS

- To minimize fluctuations in temperature only use the distilled water on your bench (in the wash bottle and in the glass 1 L bottle).
- The heating function of the heating magnetic stirrer must be **TURNED OFF** (as shown in Figure 1 below) and be sure that the stirrer plate is not hot before starting your experiment. Put the insulating plate (labeled I.P.) on top of the stirrer plate for added insulation.
- Start the stopwatch as soon as the solutions #A and #B are mixed. Stop the stopwatch as soon as the solution suddenly turns dark blue.
- Magnetic stirrer bar (take it with the provided tweezers) and beakers should be washed and rinsed with distilled water and wiped dry with paper towel to reuse.

General Procedure

Solution # A (containing $\text{Na}_2\text{S}_2\text{O}_3$, KI, KNO_3 and starch) is first placed in the beaker and is stirred using the magnetic bar. The rate of stirring is set at level 8 as indicated in Figure 1. Solution #B (containing $\text{Fe}(\text{NO}_3)_3$ and HNO_3) is quickly added into solution #A and *the stopwatch is simultaneously started. The time is recorded at the moment the solution suddenly turns dark blue.* The temperature of the solution is recorded using the digital thermometer.

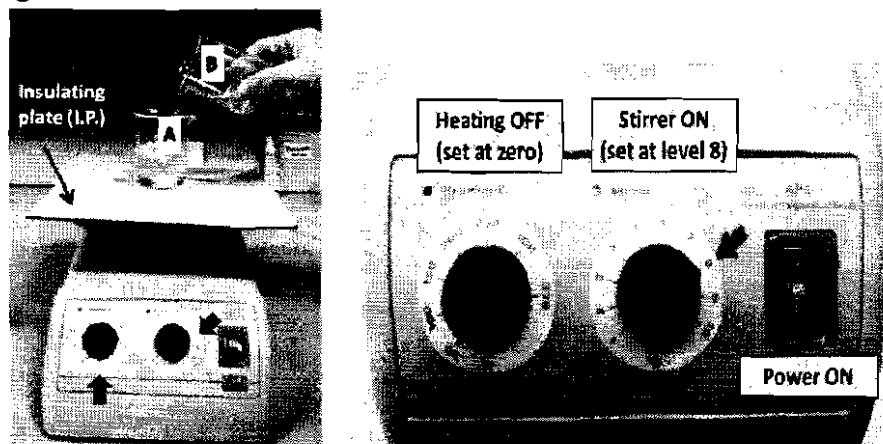


Figure 1. The apparatus employed for kinetic study of the clock reaction.



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1. Practice run to observe the color changes

- There is no need to accurately measure the volumes used in this part – just use the marks on the beaker as a guide.
- Pour *ca.* 20 mL of solution # **A1** (containing KI , $Na_2S_2O_3$, and starch in water) to a 100-mL graduated beaker containing a magnetic stirrer bar. Place the beaker on top of the insulating plate on the magnetic stirrer.
- Pour *ca.* 20 mL of solution # **B1** (containing $Fe(NO_3)_3$ and HNO_3 in water) in another 100 mL graduated beaker.
- Quickly pour the solution # **B1** into solution # **A1** and start stopwatch simultaneously. Stop stopwatch when the color of the mixture changes. There is no need to record this time. Answer the following questions.

Task 1.1: Write down the molecular formula of the limiting reactant for the given clock reaction.

$Na_2S_2O_3$ (or $S_2O_3^{2-}$)	2 points
----------------------------------	-----------------

Task 1.2: What are the ions or compounds responsible for the colors observed in this experiment? Tick the appropriate box.

Color	Compound
Purple	<input type="checkbox"/> Fe^{3+} <input type="checkbox"/> $[Fe(S_2O_3)]^+$ <input type="checkbox"/> Fe^{2+} <input type="checkbox"/> starch- I_5^- <input type="checkbox"/> I_3^-
Dark blue	<input type="checkbox"/> Fe^{3+} <input type="checkbox"/> $[Fe(S_2O_3)]^+$ <input type="checkbox"/> Fe^{2+} <input type="checkbox"/> starch- I_5^- <input type="checkbox"/> I_3^-

(4 points)

2 points for each answer

2. Determination of the order with respect to $[I^-]$ (y), and the rate constant (k)

In this section, Δt is determined for different initial concentrations of KI according to the table below. The experiment is repeated as necessary for each concentration of KI.

Hint: Use 25 mL graduated pipette for solution #A2-1, 10 mL graduated pipette for KI, 5 mL graduated pipette for solution #B2, and one of the burettes for water (you will need to refill the burette from the wash bottle for each measurement).

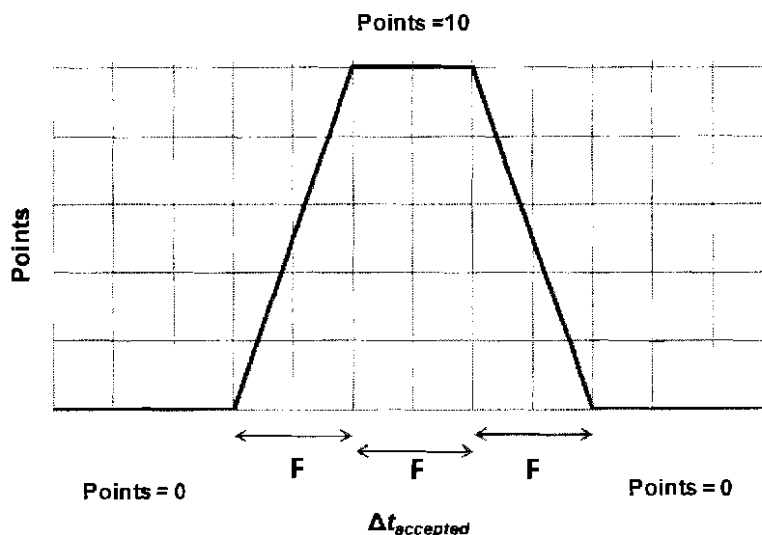
- Prepare 55 mL of solution # A2 in a 100 mL beaker containing a magnetic stirrer bar and place it on top of the insulating plate on the stirrer. Solution #A2 contains solution #A2-1, KI, and distilled water (see the table below for the volume of each component).
- Add 5 mL of solution # B2 in another 100 mL beaker.

Quickly pour prepared solution #B2 into solution #A2. Determine the time (Δt) necessary for the color change by a stopwatch. The temperature of the solution is recorded.

Task 1.3: Record the time (Δt) for each run in the table below. (You DO NOT need to fill all three columns for the runs.) For each concentration of KI, record your accepted reaction time ($\Delta t_{\text{accepted}}$) and temperature. You will be only graded on your values of $\Delta t_{\text{accepted}}$ and T_{accepted}

No	55 mL of solution #A2									$\Delta t_{\text{accepted}}$ (s)	T_{accepted} (°C)
	#A2-1 (mL)	H ₂ O (mL)	0.100M KI (mL)	Run 1		Run 2		Run 3			
				Δt (s)	T (°C)	Δt (s)	T (°C)	Δt (s)	T (°C)		
1	20.4	31.6	3.0								
2	20.4	30.1	4.5								
3	20.4	28.6	6.0								
4	20.4	27.4	7.2								
5	20.4	25.6	9.0								

09 26
 73 110
 50 152
 25 183
 20 222
 0 69



[I] (M)	T (°C)	$\Delta t_{\text{accepted}}$ (s)	$\Delta t_{\text{correct}} = m \cdot \text{EXP}(n \cdot T)$		F (s)	
			m	n		
0.0050		49.3 - 15.3	52.5	1437.0	-0.1325	6
0.0075		21.1 - 24.1	22.61	564.7	-0.1277	3
0.0100		13.37 - 15.37	14.37	323.2	-0.1276	2
0.0120		10.19 - 11.31	10.79	217.5	-0.1262	1.2
0.0150		6.56 - 7.36	6.96	114.0	-0.1170	0.8

When you are satisfied you have all the necessary data for Problem 1, before continuing further with the analysis, it is strongly recommended that you start the practical procedure for Problem 2 since there is a reaction time of one hour in that Problem.

Task 1.4: Fill in the table below and plot the results in the graph.

Hint: Make sure your data is graphed as large as possible in the provided space.

No.	1	2	3	4	5
$\ln([I]_0 / M)$	- 5.30	- 4.89	- 4.61	- 4.42	- 4.20
$\Delta t_{\text{accepted}}$ (s)					
$\ln(\Delta t_{\text{accepted}} / S)$					

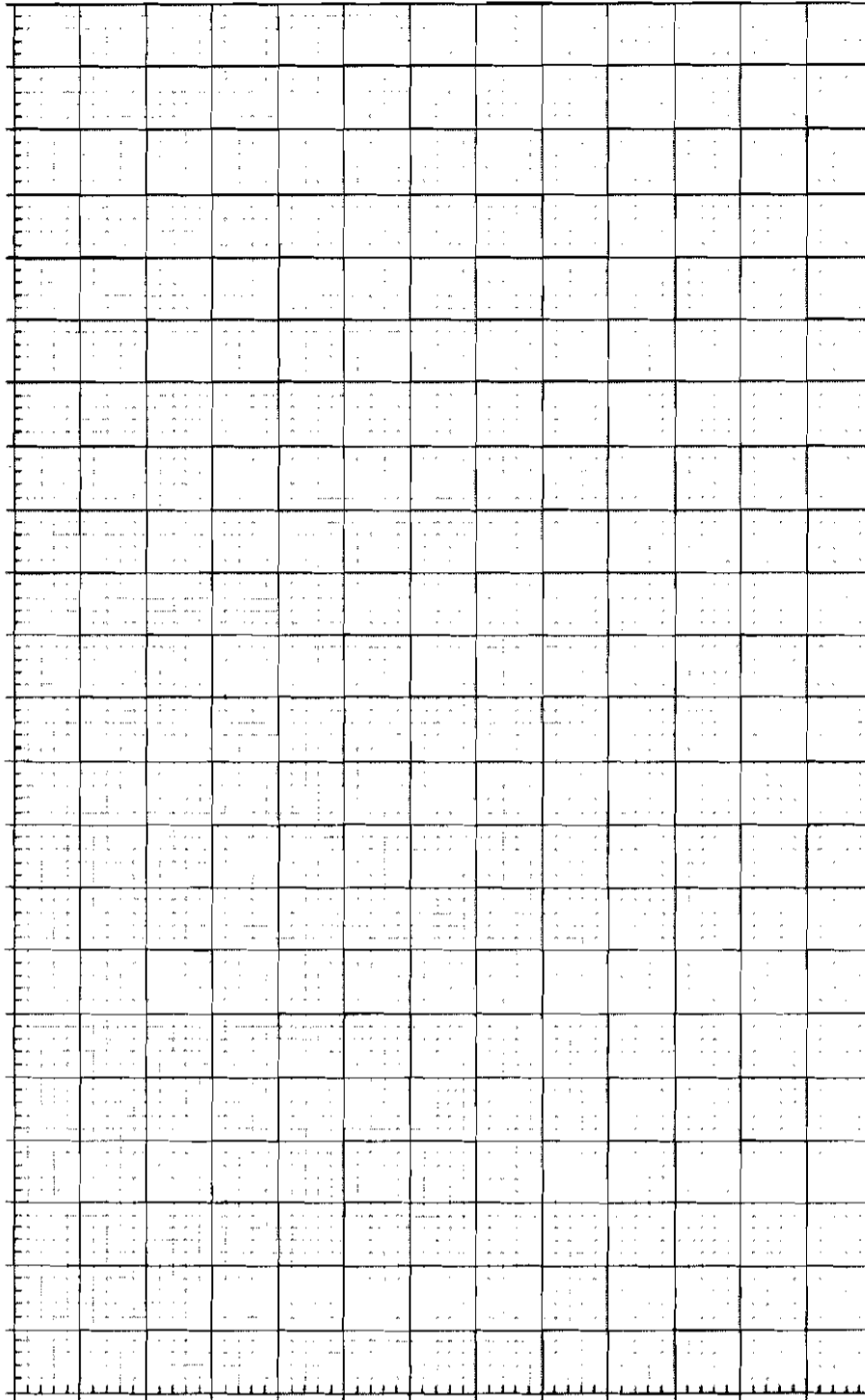


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$\ln(\Delta t_{\text{accepted}} / \text{s})$



$\ln([I^-]_0 / \text{M})$

(2 points for plotting graph)

Task 1.5: Draw the best fit line on your graph and use this to determine the order with respect to $[I^-]$ (y).

$y = 2$ 2 points for showing best fit line and reading the slope

Task 1.6: Complete the table below and calculate k for each of the concentrations of iodide. Report your accepted value for the rate constant, giving the appropriate unit. Remember that the order with respect to $[Fe^{3+}]$ is equal to one.

No	$\Delta t_{\text{accepted}}$ (s)	$[Fe^{3+}]_0$ ($\times 10^{-3}$ M)	$[I^-]_0$ ($\times 10^{-3}$ M)	$[S_2O_3^{2-}]_0$ ($\times 10^{-3}$ M)	k
1		8.5	5.0	0.2	
2		8.5	7.5	0.2	
3		8.5	10.0	0.2	
4		8.5	12.0	0.2	
5		8.5	15.0	0.2	

$k_{\text{accepted}} = \dots\dots\dots (M^{-2}s^{-1} \text{ or } mol^{-2}L^2s^{-1})$

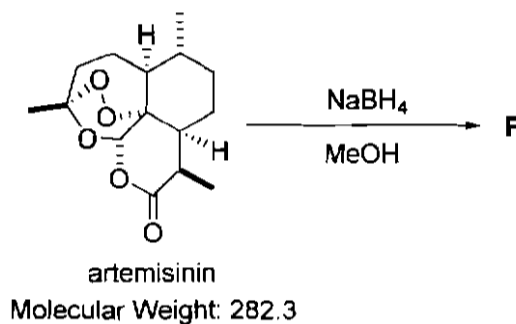
(10 points)

- +) 1 point for each correct value of k in the table.
- +) 1 point for the correct value of $[Fe^{3+}]_0$.
- +) 1 point for the correct value of $[S_2O_3^{2-}]_0$.
- +) 3 points for consistent unit of k .

Practical Problem 2 13 % of the total	Code:		Task	1	2	3	4	5	Total
	Examiner		Mark	35	15	20	4	2	76
			Grade						

Practical Problem 2. Synthesis of a derivative of Artemisinin

Artemisinin (also known as Quinghaosu) is an antimalarial drug isolated from the yellow flower herb *Artemisia annua* L., in Vietnam. This drug is highly efficacious against the chloroquine-resistant *Plasmodium falciparum*. However, artemisinin has a poor solubility in both oil and water so that one needs to prepare its new derivatives to improve the applicability of this drug. The reduction of artemisinin is an attractive method to synthesize new derivatives of artemisinin as shown in Scheme 1.

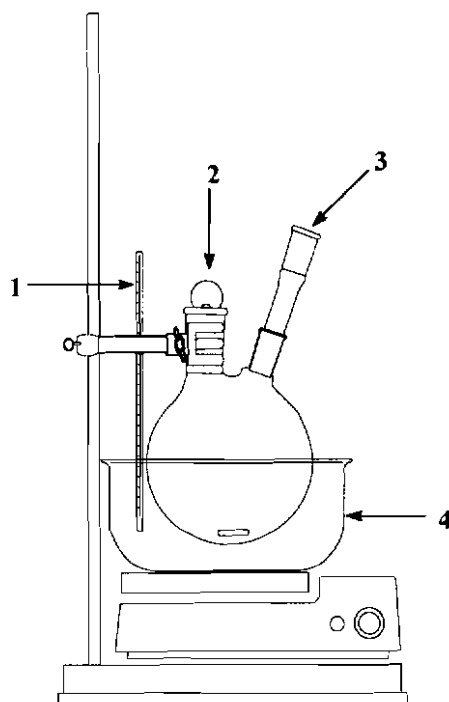


Scheme 1

In this practical exam you are going to reduce artemisinin to product **P** and check its purity using Thin-Layer Chromatography (TLC).

Experimental Set-up

- The experimental set-up is shown in Figure 2.1.
- By moving the finger clamp, you can adjust the position of the two-neck round-bottom flask.



1: Digital thermometer; 2: Plastic Stopper; 3: CaCl₂ drying tube; 4: Ice Bath

Figure 2.1. Reaction system for Problem 2

Procedure

Step 1. Synthesis of a Derivative of Artemisinin

1. Prepare an ice bath with a temperature between -20 and -15 °C by mixing ice and sodium chloride in the plastic pot (approximate ratio of NaCl : crushed ice = 1 scoop : 3 scoops). Use the digital thermometer to monitor the temperature. Place the bath on the magnetic stirrer. Put a layer of three tissues between the bath and the stirrer.
2. Connect the CaCl₂ drying tube to the small neck of the round-bottom flask and close the other neck with the plastic stopper.
3. Place a magnetic stirring bar into the dry round-bottom flask and set up the reaction system onto the clamp-stand so that the system is immersed in the ice bath. Monitor the temperature using the digital thermometer.
4. *Setting aside a tiny amount (ca. 2 mg) of artemisinin for TLC analysis*, open the stopper and add the 1 gram of artemisinin through the bigger neck.
5. Use the glass funnel to add 15 mL of methanol (measured using the 50-mL graduated cylinder). Close the stopper and turn on the magnetic stirrer. (*Set the magnetic stirrer to level 4*). Start the stopwatch to keep track of the time.
6. After *ca.* 5 min stirring, open the stopper and add carefully 0.53 g of NaBH₄ in small portions over 15 min using a spatula. Close the stopper in between addition. (*Caution: Adding NaBH₄ rapidly causes side-reactions and overflowing*). Keep stirring for 50 min. Maintain the temperature of the ice bath below -5 °C; remove



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some of the liquid and add more NaCl-crushed ice mixture if necessary. Cool the vial containing the 1 mL of acetic acid in the ice bath.

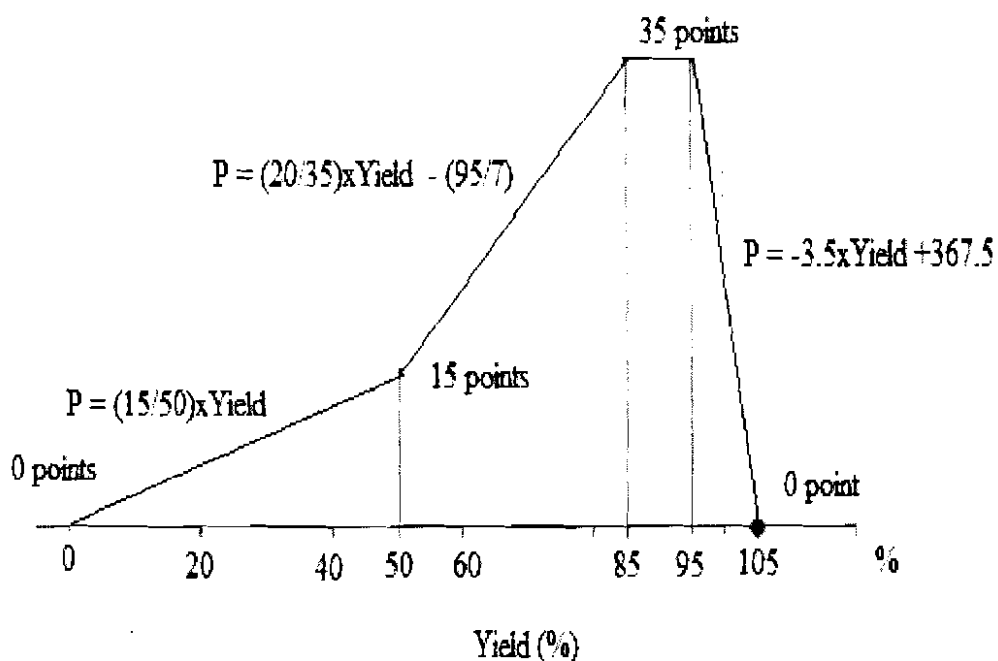
During this waiting time, you are advised to finish calculations from Problem 1, answer the questions below, and prepare further experimental steps.

7. Prepare 50 mL of ice-cold distilled water (*cooled in the ice bath*) in the 100 mL-conical flask. Measure *ca.* 20-22 mL *n*-hexane in the 50 mL measuring cylinder and cool it in the ice bath. After the reaction is complete, keep the reaction flask in the ice bath below 0 °C. Remove the CaCl₂ tube, open the stopper, and add gradually *ca.* 0.5 mL of the cold acetic acid from the vial into the reaction flask until the pH is between 6 and 7. (Use the glass rod to spot the reaction mixture on to the pH paper.) With stirring, slowly add the 50 mL of ice cold water over 2 min. A white solid precipitates in the reaction flask.
8. Assemble the vacuum filtration apparatus. Put a filter paper onto the Büchner funnel, wet the filter paper with distilled water and open the vacuum valve. Transfer the reaction mixture on to the filter, and remove the stirring bar from the reaction flask using the spatula. Wash the product three times with portions of 10 mL ice-cold water (*cooled in the ice bath*). Wash the product two times with portions of 10 mL ice-cold *n*-hexane (*cooled in the ice bath*). Continue to use the pump to dry the solid on the filter. After *ca.* 5 min, carefully transfer the dried powder on to the watch glass labeled with your code and put into the labeled Petri dish. **Turn off the vacuum valve when you do not use it!** *Note: Your sample will be collected, dried and weighed later by the lab assistant.*

Task 2.1 – the recording of your yield – will be performed after the exam by lab assistants.

Calculation of Y (%)

$$\text{Yield}(\%) = 99.28 \frac{\Delta m}{m_0}$$



Max. 35 points

Step 2. TLC Analysis of the product

1. Check your TLC plate before use. Unused damaged plates will be replaced upon request without penalty. Use the pencil to draw the start front line, and the line where the solvent front will be run to **exactly as shown in Figure 2.2**. Write your student code on the top of the TLC plate in pencil.

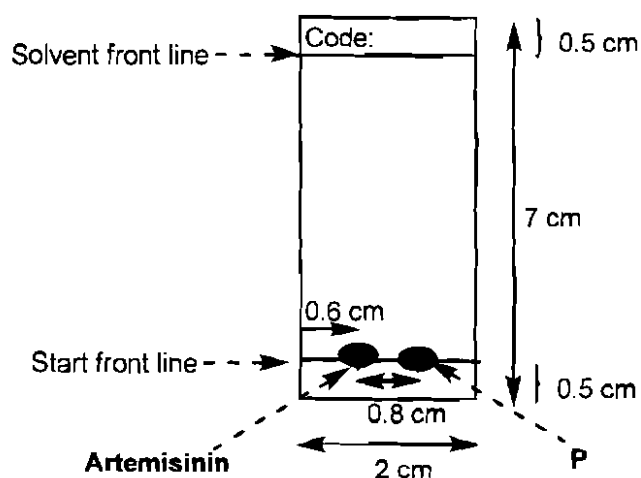


Figure 2.2. Instruction of TLC plate preparation

- Dissolve *ca.* 1 mg of artemisinin (*a spatula tip*) in *ca.* 0.5 mL of methanol in the labeled very small test tube (use the labeled 5 mL graduated pipette). Dissolve *ca.* 1 mg of the product in *ca.* 1 mL of methanol in the labeled test tube.
- Spot the artemisinin solution and the product solution on the TLC plate using two different glass capillary spotters so the finished plate is as shown in Figure 2.2.
- Prepare the TLC developing chamber. Use the 5 mL graduated cylinder to make 5 mL of a mixture of *n*-hexane/ethyl acetate (7/3, v/v) as the solvent system. Pour the mixture of *n*-hexane/ethyl acetate into the chamber (*Note: The solvent level should not reach the spots on the plate if prepared as shown*). Cover and swirl the chamber and allow it to stand for 2 min.

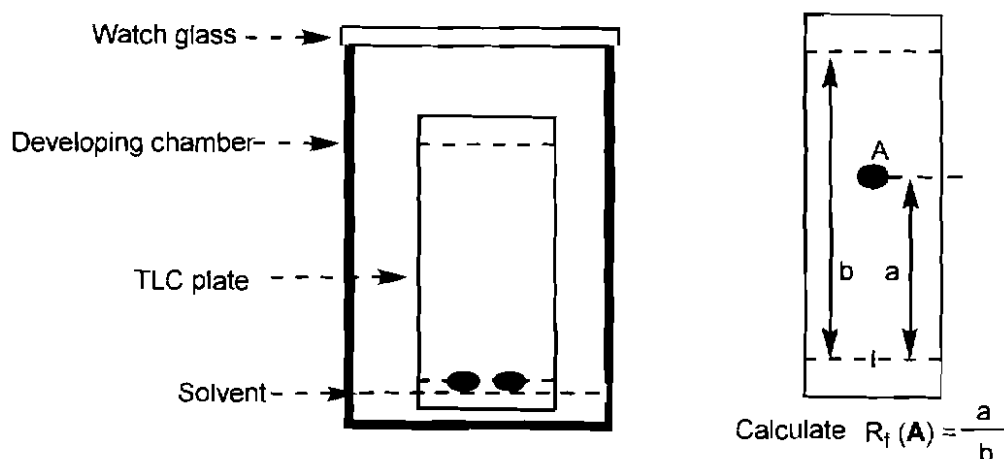


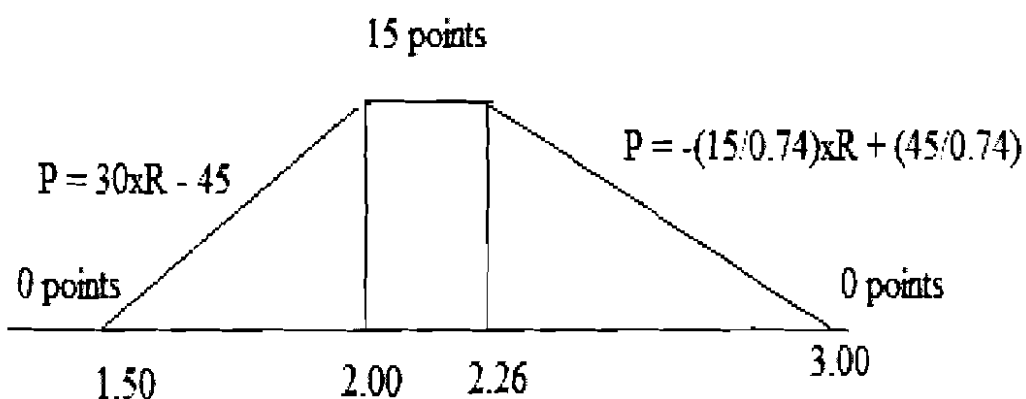
Figure 2.3. A TLC plate placed in the TLC developing chamber and instruction for R_f calculation of compound A

- Insert the TLC plate upright into the TLC developing chamber. Wait until the solvent system reaches the pre-drawn solvent front line. (*Note: You are advised to work on some question below while you wait for the TLC to run.*)
- When the solvent front reaches the line, remove the TLC plate using the tweezers and then dry the solvent using the hair dryer set at level 1.
- Dip the piece of cotton wool into the cerium staining reagent, *taking care not to let the tweezers come into contact with the solution since the metal stains the plate*. Carefully apply the stain to the whole TLC plate.
- Heat the TLC plate using the hair dryer set at level 2 (***Attention: Do NOT set the hair dryer to COLD***) until the blue spots of artemisinin and the product appear on the TLC plate.
- Ask the lab assistant to take a photo of your final TLC plate together with your student code.

10. Circle all the visualized spots and calculate the R_f values of both artemisinin and the product (See instruction in Fig. 2.3). Store your TLC plate in the Petri dish.

Task 2.2: Fill the values of R_f in Table below.

R_f , Artemisinin	R_f , Product	R_f Artemisinin/ R_f Product
-----	-----	-----
0.56 - 0.59	0.26 - 0.28	2.00 - 2.26
		15 points



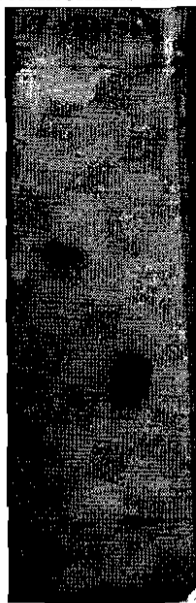
Task 2.3: Check the total number of developed spots on the TLC plate:

1	<input type="checkbox"/>	2	<input type="checkbox"/>	3	<input type="checkbox"/>	4	<input type="checkbox"/>	5	<input type="checkbox"/>
---	--------------------------	---	--------------------------	---	--------------------------	---	--------------------------	---	--------------------------

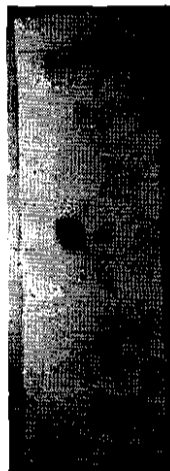
- A penalty of 5 points will be deducted from each case below if the plate is very badly run

e.g. too much sample. (No overall negative mark)

- 2 developed spots, the higher one is artemisinin, the lower one is the product, **20 points**



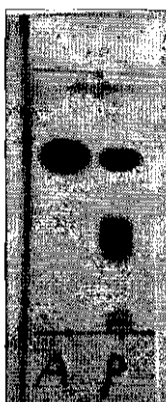
- 3 developed spots, 1 is for artemisinin, 1 is for a little amount of starting material, 1 is for product, **15 points**



- 3 developed spots, 1 is for artemisinin, 1 is for a significant amount of starting material, 1 is for product, **10 points**



- 4 developed spots, 1 is for artemisinin, 1 is for starting material, 1 for product, 1 for byproduct **5 points**



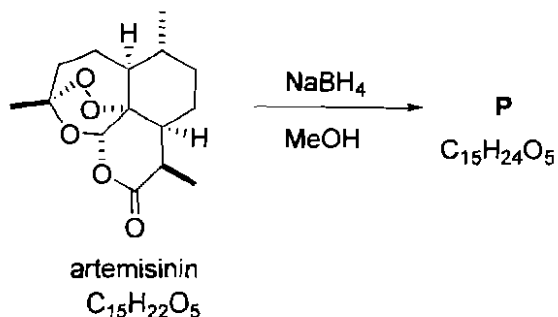
- 2 developed spots, 1 is for artemisinin, 1 is for starting material, R_f is the same, **0 point**
(also award 0 points for yield since no product has been prepared)



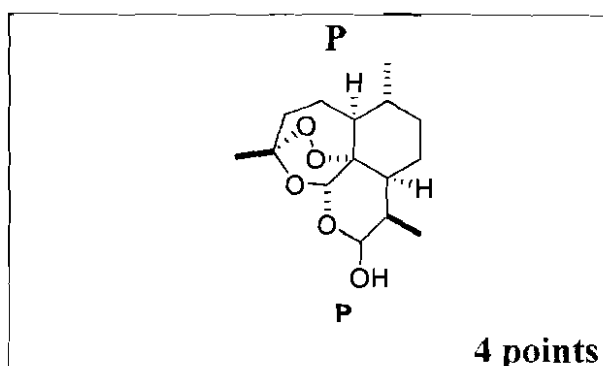
If the product is not run on TLC but the starting material has been run well, 5 points.

Step 3. Identifying the reaction product P

The reduction of artemisinin leads to the formation of two stereoisomers (**P**). Comparing the ¹H-NMR spectrum (in CDCl₃) of one of these isomers with the spectrum of artemisinin shows an extra signal at δ_H = 5.29 ppm as a doublet, and also an extra signal as a broad singlet at δ_H = 2.82 ppm.



Task 2.4: Suggest structure for **P**. (You do not need to draw the stereochemistry of the compounds).



Task 2.5: **P** is mixture of two stereoisomers. What is their stereochemical relationship? Check the appropriate box below.

Z/E Isomers	Enantiomers	Diastereomers	Constitutional Isomers
<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>

2 points



Name:

Ser #2

Code:

0, 25, 4, 99, 9, 10

Signature:

Practical Problem 3 13 % of the total	Code:		Task	1	2	3	4	5	6	7	8	9	10	Total
	Examiner		Mark	0	25	2	25	3	4	3	2	5	2	71
			Grade											

Practical Problem 3. Analysis of a hydrated zinc iron(II) oxalate double salt

Zinc iron(II) oxalate double salt is a common precursor in the synthesis of zinc ferrite which is widely used in many types of electronic devices due to its interesting magnetic properties. However, such double salts may exist with different compositions and different amount of water depending on how the sample was synthesized.

You will analyze a pure sample of hydrated zinc iron(II) oxalate double salt (**Z**) in order to determine its empirical formula.

Procedure

The concentration of the standard KMnO_4 is posted on the lab walls.

Bring a clean 250 mL beaker to the lab assistant who will be waiting by the balance. You will receive a pure sample of **Z** for analysis. Accurately weigh between 0.7-0.8 g of the pure sample **Z** onto the weighing paper (m , grams). This should then be immediately quantitatively transferred into your 250 mL beaker for analysis, and its mass recorded in table below.

Task 3.1: Record the mass of the sample of pure **Z taken.**

Mass of sample, m (gram)	Lab assistant's signature
-----	-----

0 points

Analysis of **Z**

- Using the 100 mL graduated measuring cylinder, measure *ca.* 30 mL of 30 wt% H₂SO₄ solution and add it into the 250-mL beaker containing your accurately weighed pure sample of **Z**. To speed up the dissolving of your sample you may use the hotplate stirrer to warm up the mixture, **but be careful not to boil it**. *You should not use the digital thermometer as the acid may damage it.* After the solid has dissolved, remove the beaker from the hotplate stirrer and cool it to close to room temperature. After the solution has cooled, quantitatively transfer it into the 100 mL volumetric flask. Add distilled water up to the 100 mL–mark. We will now call this solution **C**.
- Use an appropriately labeled beaker to transfer the standardized **KMnO₄** solution into the burette graduated with **brown** marks.
- Use another appropriately labeled beaker to transfer the standardize **EDTA** solution into the burette graduated with **blue** marks.

Titration with KMnO₄

- a) Using the 5 mL graduated pipette add 5.00 mL of the solution **C** into a 250 mL conical flask.
- b) To this conical flask add about 2 mL of 30 wt% H₂SO₄ solution, about 3 mL of 3.0 M H₃PO₄ solution, and about 10 mL of distilled water. Heat the mixture on the hot plate stirrer until hot, **but be careful not to boil it**.
- c) Titrate the hot solution with the standardized KMnO₄ solution, recording your burette readings in the table below. At the end point of the titration, the pink color of the solution appears. Repeat the titration as desired and report your accepted volume of KMnO₄ solution consumed (V₁ mL) in the table.

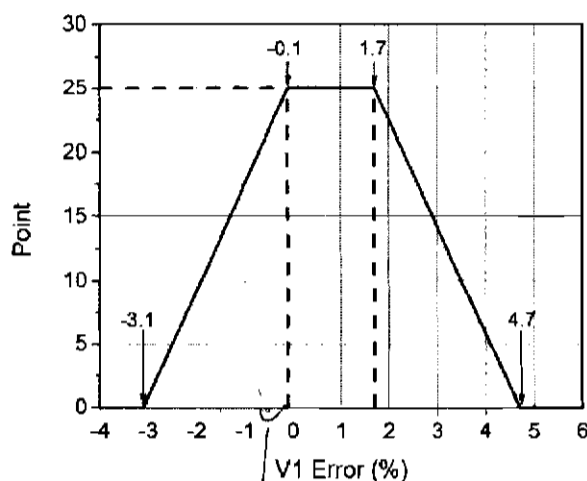
Task 3.2: Record volumes of standardized KMnO₄ solution consumed

(You DO NOT need to fill in the entire table)

	Titration No			
	1	2	3	4
Initial reading of the burette of KMnO ₄ , mL				
Final reading of the burette of KMnO ₄ , mL				
Consumed volume of KMnO ₄ , mL				

Accepted volume, $V_1 = \underline{\hspace{2cm}}$ mL

If V_1 error < -3.1 and V_1 error > 4.7 then $P = 0$
 If $-3.1 \leq V_2$ error < -0.1 then $P = 8.3333 \times V_1$ error + 25.8333
 If $-0.1 \leq V_2$ error < 1.7 then $P_{\max} = 25$ points
 If $1.7 \leq V_2$ error < 4.7 then $P = -8.3333 \times V_1$ error + 39.1667



Task 3.3: Can aqueous HCl or HNO₃ be used instead of H₂SO₄ for the dissolving of sample Z and the subsequent analyses?

HCl	YES <input type="checkbox"/>	NO <input checked="" type="checkbox"/>	1 pt
HNO ₃	YES <input type="checkbox"/>	NO <input checked="" type="checkbox"/>	1 pt

2.1 Titration with EDTA

- Clean both the 250 mL beakers ready for the next part of the experiment. Pipette 10.00 mL of solution C into a 250 mL beaker. Heat and stir the solution on the hotplate stirrer, **but be careful not to boil it**. Add *ca.* 15 mL of 20 wt% NaOH solution to the beaker and keep it on the hotplate for *ca.* 3-5 min in order to



Name:

Code:

Signature:

complete the precipitation of iron hydroxide, and to convert all Zn^{2+} ions into the ionic complex $[\text{Zn}(\text{OH})_4]^{2-}$.

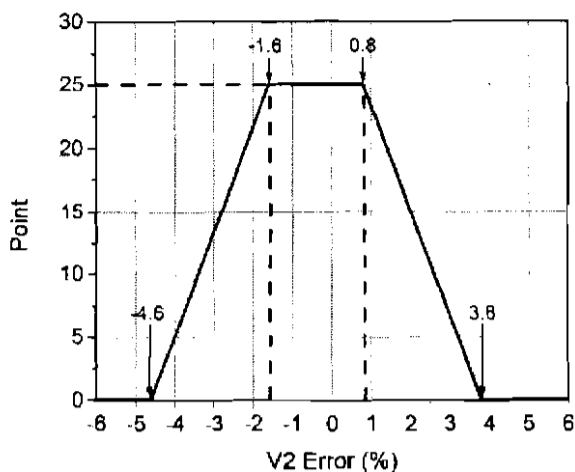
- Using a glass funnel and the large quantitative filter paper, filter the hot suspension directly into the 250 mL conical flask. **From this point take care with the volumes as you will be preparing a standard solution of exactly 100 mL from the filtrate.** As it is filtering, prepare some warm distilled water in a 250 mL beaker (*ca.* 50 mL). Wash the precipitate on the filter paper (at least 5 times) with small portions (*ca.* 5 mL) of the warm distilled water. Cool the filtrate down and then quantitatively transfer it into the 100 mL volumetric flask via a glass funnel. Add distilled water to make up to the 100 mL mark. This will now be referred to as solution **D**.
- Pipette 10.00 mL of solution **D** into a 250 mL conical flask. Add *ca.* 10 mL ammonia buffer solution ($\text{pH} = 9 - 10$) and a small quantity of the ETOO indicator using the glass spatula spoon. Mix well to obtain a purple solution. Titrate the solution with the standardized 2.00×10^{-3} M EDTA solution, recording your burette readings in table below. At the end point, the color of the solution turns blue. Repeat the titration as desired and report your accepted volume of EDTA solution consumed (V_2 mL) in the table.

Task 3.4: Record the volumes of EDTA solution consumed

(You DO NOT need to fill in the entire table)

	Titration No			
	1	2	3	4
Initial reading of the burette of EDTA, mL				
Final reading of the burette of EDTA, mL				
Consumed volume of EDTA, mL				

Accepted volume, $V_2 = \underline{\hspace{2cm}} \text{ mL}$

$V_2, \text{ mL} = \dots\dots\dots$
25 pts


- If V_2 error < -4.6 and V_2 error > 3.8 then $P = 0$
- If $-4.6 \leq V_2$ error < -1.6 then $P = 8.333 \times V_2$ error $+ 38.3333$
- If $-1.6 \leq V_2$ error < 0.8 then $P_{\text{max}} = 25$ points
- If $0.8 \leq V_2$ error < 3.8 then $P = -8.333 \times V_2$ error $+ 31.6667$

Determination of the empirical formula of Z

Task 3.5: Calculate the number of moles of Zn^{2+} , $n_{\text{Zn}^{2+}}$, present in 100 mL of solution

C.

0.1 M x 1 M EDTA
 $M_{\text{EDTA}} \times V_{\text{EDTA}} = 0.1 M \times V_{\text{Zn}^{2+}}$

The obtained value of $n_{\text{Zn}^{2+}}$ (mol): 3pts

Task 3.6: Give the ionic equations for the reduction-oxidation reactions taking place in the titration with KMnO_4 .

$5\text{Fe}^{2+} + \text{MnO}_4^- + 8\text{H}^+ \rightarrow 5\text{Fe}^{3+} + \text{Mn}^{2+} + 4\text{H}_2\text{O}$ 2 pts

$5\text{C}_2\text{O}_4^{2-} + 2\text{MnO}_4^- + 16\text{H}^+ \rightarrow 10\text{CO}_2 + 2\text{Mn}^{2+} + 8\text{H}_2\text{O}$ 2 pts

(or $5\text{H}_2\text{C}_2\text{O}_4 + 2\text{MnO}_4^- + 6\text{H}^+ \rightarrow 10\text{CO}_2 + 2\text{Mn}^{2+} + 8\text{H}_2\text{O}$)

Task 3.7: Calculate the number of moles of Fe^{2+} , $n_{Fe^{2+}}$, present in 100 mL of solution

C. [YOU WILL NEED THE PRECISE CONCENTRATION OF $KMnO_4$ POSTED ON THE WALLS IN YOUR LAB]

V_1 , mL =

$$n_{Fe^{2+}} = \frac{5 \times 20 \times V_1 \times C_{KMnO_4} - 2n_{Zn^{2+}}}{3}$$

$n_{Fe^{2+}}$ (mol): 3 pts

Task 3.8: Calculate the number of moles of $C_2O_4^{2-}$ anion, $n_{C_2O_4^{2-}}$, in 100 mL of solution C.

$$n_{C_2O_4^{2-}} = n_{Zn^{2+}} + n_{Fe^{2+}}$$

Calculate the molar number of $C_2O_4^{2-}$ anions in solution C (mole): ----- 2 pts

Task 3.9: Calculate the number of moles of water, n_{H_2O} , in the original sample of Z taken for analysis.

5 points

Task 3.10: Give the empirical formula of Z:

$ZnFe_2(C_2O_4)_3 \cdot 6H_2O$

2 pts

Chemistry: The flavor of life

Periodic Table of the Elements

1	1 H 1.008	2	Transition Elements										13	14	15	16	17	18 2 He 4.003
2	3 Li 6.941	4 Be 9.012											5 B 10.81	6 C 12.01	7 N 14.01	8 O 16.00	9 F 19.00	10 Ne 20.18
3	11 Na 22.99	12 Mg 24.31	3	4	5	6	7	8	9	10	11	12	13 Al 26.98	14 Si 28.09	15 P 30.98	16 S 32.07	17 Cl 35.45	18 Ar 39.95
4	19 K 39.10	20 Ca 40.08	21 Sc 44.96	22 Ti 47.87	23 V 50.94	24 Cr 52.00	25 Mn 54.94	26 Fe 55.85	27 Co 58.93	28 Ni 58.69	29 Cu 63.55	30 Zn 65.41	31 Ga 69.72	32 Ge 72.61	33 As 74.92	34 Se 78.96	35 Br 79.90	36 Kr 83.80
5	37 Rb 85.47	38 Sr 87.62	39 Y 88.91	40 Zr 91.22	41 Nb 92.91	42 Mo 95.94	43 Tc (97.9)	44 Ru 101.1	45 Rh 102.9	46 Pd 106.4	47 Ag 107.9	48 Cd 112.4	49 In 114.8	50 Sn 118.7	51 Sb 121.8	52 Te 127.6	53 I 126.9	54 Xe 131.3
6	55 Cs 132.9	56 Ba 137.3	57 La 138.9	72 Hf 178.5	73 Ta 180.9	74 W 183.8	75 Re 186.2	76 Os 190.2	77 Ir 192.2	78 Pt 195.1	79 Au 197.0	80 Hg 200.6	81 Tl 204.4	82 Pb 207.2	83 Bi 209.0	84 Po (209.0)	85 At (210.0)	86 Rn (222.0)
7	87 Fr (223.0)	88 Ra	89 Ac (227.0)	104 Rf (261.1)	105 Db (262.1)	106 Sg (263.1)	107 Bh (262.1)	108 Hs (265)	109 Mt (266)	110 Ds (271)	111 Rg (272)	112 Cn (285)	113 Uut (284)	114 Fl (289)	115 Uup (288)	116 Lv (292)	117 Uus (294)	118 Uuo (294)

6	Lanthanides	58 Ce 140.1	59 Pr 140.9	60 Nd 144.2	61 Pm (144.9)	62 Sm 150.4	63 Eu 152.0	64 Gd 157.3	65 Tb 158.9	66 Dy 162.5	67 Ho 164.9	68 Er 167.3	69 Tm 168.9	70 Yb 173.0	71 Lu 174.0
7	Actinides	90 Th 232.0	91 Pa 231.0	92 U 238.0	93 Np (237.1)	94 Pu (244.1)	95 Am (243.1)	96 Cm (247.1)	97 Bk (247.1)	98 Cf (251.1)	99 Es (252.1)	100 Fm (257.1)	101 Md (258.1)	102 No (259.1)	103 Lr (260.1)